

A22

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization  
International Bureau



(43) International Publication Date  
8 November 2001 (08.11.2001)

PCT

(10) International Publication Number  
**WO 01/83799 A1**

(51) International Patent Classification<sup>7</sup>: **C12P 13/02**

(DE). **HERMANN, Thomas**; Zirkonstrasse 8, 33739 Bielefeld (DE).

(21) International Application Number: **PCT/EP01/04651**

(22) International Filing Date: **25 April 2001 (25.04.2001)**

(25) Filing Language: **English**

(26) Publication Language: **English**

(30) Priority Data:  
**100 21 515.7** **3 May 2000 (03.05.2000)** **DE**

(71) Applicant: **DEGUSSA AG** [DE/DE]; Bennigsenplatz 1, 40474 Düsseldorf (DE).

(72) Inventors: **PFEFFERLE, Walter**; Jahnstrasse 33, 33790 Halle (Westf.) (DE). **BECKER, Ulrich**; Prieckhof 318, 97611 Selze (SK). **WALGER, Ilona**; Barlachstrasse 7, 33607 Bielefeld (DE). **BINDER, Michael**; Kalberkamp 28, 33803 Steinhagen (Westf.) (DE). **UFFMANN, Klaus-Erich**; Rappoldstrasse 73, 33611 Bielefeld (DE). **THIERBACH, Georg**; Gunststrasse 21, 33613 Bielefeld (DE). **MARX, Achim**; Altenbrede 4B, 33613 Bielefeld

(81) Designated States (*national*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.

(84) Designated States (*regional*): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

— with international search report

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



**WO 01/83799 A1**

(54) Title: **PROCESS FOR THE PREPARATION OF ALKALINE EARTH SALTS OF D-PANTOTHENIC ACID**

(57) Abstract: The invention relates to the preparation of alkaline earth salts of D-pantothenic acid, in which the fermentation is carried out in the presence of alkaline earth compounds.

**Process for the Preparation of Alkaline Earth Salts of  
D-Pantothenic Acid**

The present invention relates to a process for the preparation of alkaline earth salts of D-pantothenic acid  
5 from fermentation broths.

Prior art.

Pantothenic acid is a commercially important vitamin that is used in cosmetics, medicine, human nutrition and in animal nutrition.

10 Pantothenic acid can be prepared by chemical synthesis or via biotechnology by fermentation of suitable microorganisms in suitable nutrient solutions. DL-pantolactone is an important compound in the chemical synthesis, and is prepared in a multi-stage process from  
15 formaldehyde, isobutyl aldehyde and cyanide. In further process steps the racemic mixture is separated and D-pantolactone is condensed with  $\beta$ -alanine to yield D-pantothenic acid.

The advantage of a fermentative preparation using  
20 microorganisms is the direct formation of the correct stereoisometric form, namely the D-form of pantothenic acid free from L-pantothenic acid.

Various types of bacteria, for example *Escherichia coli*, *Arthrobacter ureafaciens*, *Corynebacterium erythrogenes*,  
25 *Brevibacterium ammoniagenes* and also yeasts, for example *Debaromyces castellii* may, as demonstrated in EP-A-0 493 060, EP-A-0 590 857 and WO 97/10340, produce D-pantothenic acid under suitable fermentation conditions. Particularly suitable microorganisms are the derivatives, described in  
30 the citations, of *Escherichia coli* IF03547, for example the strains FV5069/pFV31 or FV5069/pFV202.

In the fermentative preparation of D-pantothenic acid as is described in EP-A-0 493 060, EP-A-0 590 857 and WO 97/10340, a microorganism capable of producing D-pantothenic acid is cultivated in a suitable nutrient medium and the D-pantothenic acid that is formed is then isolated according to a complicated expensive procedure, purified, and obtained as the calcium salt.

Suitable nutrient media contain a carbon source such as glucose or starch flour hydrolysate, precursors such as  $\beta$ -alanine, D,L-pantoic acid or D,L-pantolactone, a nitrogen source such as ammonium sulfate, a phosphorus source such as potassium phosphate, and further salts, trace elements, amino acids and vitamins, and optionally complex media additives such as yeast extract or corn steep liquor. The microorganisms are then incubated in this medium at a suitable pH value under appropriate aeration and stirring, whereupon the microorganisms form D-pantothenic acid.

EP-A-0 590 857 describes for example a fed batch process for preparing pantothenic acid in a 5 l reactor filled with 2.3 l or 2.5 l of culture medium. In this experimental example solid calcium carbonate was added presumably to regulate the pH. The preliminary addition or subsequent addition of solid calcium carbonate is however extremely undesirable in a large-scale reactor having a volume of many cubic metres, because the material loading is increased by the calcium carbonate deposits on the stirrer blades, internal surfaces and seals, and the flow properties of the culture liquid and the sterile conditions are adversely affected.

According to the present prior art, which is outlined in WO96/33283 and EP-A-0 590857, the calcium salt of D-pantothenic acid is obtained by a complicated and costly isolation and purification process starting from a fermentation broth containing pantothenic acid. After a

first separation of the biomass by filtration or centrifugation, the filtrate is worked up further by purification by means of activated charcoal or by column chromatography. After adding calcium hydroxide to the  
5 pretreated filtrate or eluate, the batch is then purified by crystallisation.

The purification method described in WO96/33283 is carried out as follows. The filtrate is decolourised by means of activated charcoal in a first column. The pH is adjusted  
10 to 3.0 with concentrated hydrochloric acid and the liquid is then purified continuously through two columns packed with activated charcoal. The elution of the D-pantothenic acid is performed with methyl alcohol. A subsequent neutralisation is carried out with  $\text{Ca}(\text{OH})_2$  powder while  
15 thoroughly mixing. The calcium D-pantothenate is obtained by subsequent crystallisation at 5°C.

The purification method described in EP-A 0 590 857 is carried out as follows. The filtrate is first of all purified with the aid of cation exchange and anion exchange  
20 columns. Elution is performed with hydrochloric acid. The eluted fraction is then neutralised with  $\text{Ca}(\text{OH})_2$ , following which activated charcoal is added and the mixture is filtered. The resultant filtrate is extracted in a low molecular weight alcohol (methanol, ethanol, isopropanol)  
25 and the calcium D-pantothenate is obtained by crystallisation.

The calcium D-pantothenate prepared in the aforescribed manner is used as a feed additive for animal nutrition.

Object of the invention

30 The inventors have provided an improved process for preparing alkaline earth salts, in particular the calcium and magnesium salts, of D-pantothenic acid, which are suitable for use as feed additives in animal nutrition.

### Description of the invention

The vitamin D-pantothenic acid is a commercially important product that is used in animal nutrition, medicine, human nutrition and in cosmetics. There is therefore a general  
5 interest in providing new processes for preparing pantothenic acid or its salts.

The present invention provides a process for preparing alkaline earth salts of D-pantothenic acid or mixtures containing the latter, from fermentation broths, which is  
10 characterised in that

- a) the fermentation is carried out in the presence of alkaline earth compound,
- b) after completion of the fermentation the biomass is optionally removed either in whole or in part,
- 15 c) the thus worked up fermentation broth is concentrated, and
- d) the alkaline earth salt or salts of D-pantothenic acid is/are obtained from the latter in pure form or as a mixture containing the constituents of the fermentation  
20 broth.

The present invention also provides a process which is characterised in that one or more various alkaline earth salt(s) of D-pantothenic acid is/are added in the desired amount to the constituents of the fermentation broth and  
25 mixture containing one or more of the alkaline earth salts of D-pantothenic acid.

The invention furthermore provides a process for preparing alkaline earth salts of D-pantothenic acid, characterised in that

- a) the biomass is separated from a fermentation broth containing alkaline earth salt(s) of D-Pantothenic acid,
  - b) the cell-free fermentation broth is concentrated,
  - 5 c) a hydrophilic organic solvent, in particular ethanol, methanol or acetone, is added to the concentrate thus obtained, following which
  - d) the alkaline earth salt(s) of pantothenic acid is/are isolated, optionally washed with a hydrophilic organic  
10 solvent and then, if desired,
  - e) recrystallised in an aqueous solution of a hydrophilic organic solvent, optionally with the addition of activated charcoal, and obtained in a high state of purity.
- 15 All microorganisms that are capable of producing D-pantothenic acid and which produce D-pantothenic acid under appropriate fermentation conditions are suitable for the process according to the invention. The microorganisms can produce pantothenic acid from glucose, sucrose, lactose,  
20 fructose, maltose, molasses, starch, cellulose or from glycerol and ethanol.

The microorganisms may be fungi or yeasts, for example *Debaromyces castellii* or *Saccharomyces cerevisiae* or Gram-positive bacteria, for example of the genus *Corynebacterium*  
25 or may be Gram-negative bacteria, for example of the family of Enterobacteriaceae. Within the family of Enterobacteriaceae, the genus *Escherichia* as exemplified by the type *Escherichia coli* should in particular be mentioned. Within the type *Escherichia coli* there should  
30 be mentioned the so-called K-12 strains, for example the strains MG1655 or W3110 (Neidhard et al.: *Escherichia coli* and *Salmonella*. Cellular and Molecular Biology (ASM Press,

Washington D.C.)) or the *Escherichia coli* wild type strain IFO3547 (Institute for Fermentation, Osaka, Japan) and mutants derived therefrom. Among the strains obtained from IFO3547, there should in turn be mentioned FV5069/pFV31  
5 (EP-A-0 590 857, US-A 5 518 906) and FV5069/pFV202 (WO 97/10340, US-A-5 932 457). In the genus *Corynebacterium* the type *Corynebacterium glutamicum* should in particular be mentioned.

The aforescribed microorganisms may be cultivated  
10 continuously or discontinuously in a batch process or in a fed batch or repeated fed batch process in order to produce alkaline earth salts of D-pantothenic acid. A summary of known cultivation methods is described in the handbook by Chmiel (Bioprozesstechnik 1. Einführung in die  
15 Bioverfahrenstechnik [Introduction to Biotechnology] (Gustav Fischer Verlag, Stuttgart, 1991)) or in the handbook by Storhas (Bioreaktoren und periphere Einrichtungen (Vieweg Verlag, Brunswick/Wiesbaden, 1994)).

The culture medium to be used must suitably satisfy the  
20 requirements of the relevant microorganisms. Descriptions of culture media for various microorganisms are contained in the handbook "Manual of Methods for General Bacteriology" of the American Society for Bacteriology (Washington D.C., USA, 1981). Sugars and carbohydrates,  
25 for example glucose, sucrose, lactose, fructose, maltose, molasses, starch and cellulose, oils and fats such as soya bean oil, sunflower oil, groundnut oil and coconut oil, fatty acids, for example palmitic acid, stearic acid and linoleic acid, alcohols such as glycerol and ethanol, and  
30 also organic acids such as acetic acid may be used as sources of carbon. These substances may be used individually or as a mixture. As nitrogen source there may for example be used organic nitrogen-containing compounds such as peptones, yeast extract, meat extract, malt  
35 extract, corn steep liquor, soya bean flour and urea, or

inorganic compounds such as ammonium sulfate, ammonium chloride, ammonium phosphate, ammonium carbonate and ammonium nitrate. The nitrogen sources may be used individually or as a mixture. As phosphorus source there  
5 may be used potassium dihydrogen phosphate or dipotassium hydrogen phosphate or the corresponding sodium-containing salts. The culture medium must also contain metal salts such as magnesium sulfate or iron sulfate that are necessary for growth. Finally, essential growth factors  
10 such as amino acids and vitamins may additionally be used together with the aforementioned substances. Precursors such as  $\beta$ -alanine or optionally their salts may furthermore be added to the culture medium. The aforementioned substances may be added to the culture in the form of a  
15 one-off addition or may be appropriately metered in during the cultivation.

Basic compounds such as ammonia or ammonia water or acidic compounds such as phosphoric acid or sulfuric acid may be used in an appropriate manner to control the pH of the  
20 culture, insofar as the process according to the invention does not involve any other measures. Anti-foaming agents such as fatty acid polyglycol esters may be used to control foam formation. Suitable selectively acting substances, for example antibiotics, may be added to maintain the  
25 stability of plasmids. Oxygen or oxygen-containing gas mixtures, for example air, are introduced into the culture in order to maintain aerobic conditions. The temperature of the culture is normally 20°C to 50°C, and preferably 25°C to 45°C. Cultivation is continued until a maximum  
30 amount of D-pantothenic acid has been formed. This objective is normally achieved within 10 hours to 160 hours.

It has been found that the alkaline earth salts, in particular the calcium salt and magnesium salt of D-  
35 pantothenic acid, can be produced in a simple manner by



adding during the fermentation a solution or suspension of an alkaline earth-containing inorganic compound, such as for example calcium hydroxide, magnesium hydroxide, in particular calcium hydroxide, or magnesium oxide, or also  
5 the alkaline earth salt of an organic acid, for example calcium acetate, calcium fumarate or calcium aspartate, continuously to the fermentation. It is also possible to meter in the alkaline earth-containing compound batchwise. In this way the cation necessary for forming the desired  
10 alkaline earth salt of D-pantothenic acid is introduced directly into the fermentation broth.

The fermentation process according to the invention is generally characterised in that a microorganism capable of producing D-pantothenic acid is first of all cultivated in  
15 a known manner using ammonia as pH regulator and nitrogen source, and in the following production stage the pH is preferably adjusted by using a solution or suspension of an alkaline earth-containing compound, for example calcium hydroxide, magnesium hydroxide, magnesium oxide, calcium  
20 acetate, calcium fumarate or calcium aspartate. If calcium salts of organic acids are used, the organic radicals are generally utilised as a carbon source by the microorganisms.

The solutions or suspensions of the alkaline earth-  
25 containing compounds that are used, in particular calcium hydroxide and magnesium hydroxide, have a concentration of 5 - 50 wt.%, preferably 5 - 30 wt.%, the range from 10 - 25 wt.% being most particularly preferred. It is also possible according to the invention to employ mixtures of  
30 various alkaline earth-containing compounds in these concentration ranges.

The alkaline earth compound is metered in in such a way that the molar ratio of alkaline earth compound to the formed D-pantothenic acid is in the range 1 : 0,5 to

1 : 20, preferably in the range 1 : 1.3 to 1 : 10, more preferably in the range 1 : 1.3 to 1 : 2.5, the stoichiometric range from 1 : 1.8 to 1 : 2.2 being most particularly preferred. If necessary, during the course of the fermentation the pH can be adjusted by metering in ammonia in aqueous form or as a gas should there be an excessive formation of byproducts containing carboxyl groups, in order to maintain the ratio of alkaline earth compound to formed D-pantothenic acid within the desired range.

The metering in of the alkaline earth compound may take place after a fermentation time of 1 - 70 hours, preferably 10 - 40 hours and most preferably 20 - 25 hours. The concentration of D-pantothenic acid at the start of the metering in of the alkaline earth compound is generally 0.5 - 70 g/l, preferably 5 - 35 g/l and particularly preferably 20 - 25 g/l. The concentration of biomass at the time the alkaline earth compound is metered in is generally 1 - 30 g dry weight/l, preferably 10 - 23 g dry weight/l and particularly preferably 17 - 21 g dry weight/l.

The present invention also provides a process for producing powders or feed additives containing alkaline earth salts, in particular the calcium or magnesium salt of D-pantothenic acid, in a quick and cost-effective manner. To this end a fermentation broth prepared according to the process of the invention and containing in particular calcium or magnesium D-pantothenate is concentrated using known methods, for example with the aid of a rotary evaporator, thin-film evaporator or falling-film evaporator. The fermentation broth that has been concentrated in this way is then converted by spray drying or freeze drying techniques, such as are described for example in the handbook by M.L. Shuler and F. Kargi "Bioprocess Engineering, Basic Concepts" (Prentice Hall Inc., Englewood Cliffs, New Jersey, USA, 1992), or by other

appropriate methods, into a preferably free-flowing powder or feed additive. A substance or preparation containing calcium or magnesium D-pantothenate may optionally be added at a suitable process stage in order to achieve a desired concentration level. The concentration of calcium or magnesium D-pantothenate - expressed as D-pantothenic acid - in the resultant product is 20 to 80 wt.%, preferably 30 to 75 wt.%. The product is suitable as a feed additive for use in animal nutrition.

10 The inventors have also found a further method of producing powders or feed additives containing calcium or magnesium D-pantothenate. To this end a fermentation broth prepared as described above and containing in particular calcium or magnesium D-pantothenate is first of all freed either

15 completely or partially from the biomass by known separation methods, for example centrifugation, filtration, decanting or a combination of the latter. The cell-free fermentation broth is then concentrated using known methods, for example with the aid of a rotary evaporator,

20 thin-film evaporator or falling-film evaporator. The suspension concentrated in this manner is then worked up by methods involving spray drying or freeze drying or by other suitable processes into a preferably free flowing powder. A substance or preparation containing calcium or magnesium

25 D-pantothenate is optionally added at a suitable process stage in order to achieve a desired concentration value. The concentration of calcium or magnesium D-pantothenate - expressed as D-pantothenic acid - in the resultant product is 20 to 80 wt.%, preferably 30 to 75 wt.%. The product is

30 suitable for use as a feed additive in animal nutrition.

The inventors have likewise discovered a method for producing crystals containing calcium or magnesium D-pantothenate in a quick and cost-effective manner. To this end a fermentation broth prepared as described above and

35 containing calcium or magnesium D-pantothenate is first of

all freed as described above from the biomass. The cell-free fermentation broth is then concentrated using known methods, for example with the aid of a rotary evaporator, thin-film evaporator or falling-film evaporator. A  
5 hydrophilic, organic solvent, for example ethanol, methanol or acetone is then added to the suspension that has been concentrated in this way. Residual solid material is precipitated out by cooling the suspension to 0 - 8°C, preferably 0 - 5°C. The desired salt is then crystallised  
10 by inoculating with crystalline calcium or magnesium D-pantothenate or substances containing crystalline calcium or magnesium D-pantothenate. The crystallisation takes place at 0 - 8°C, preferably 0 - 5°C, and lasts from 1 hour to 12 days, preferably 1 hour to 10 days, and particularly  
15 preferably 1 hour to 8 days. The crystalline crop obtained in this way is preferably washed with methanol and then dried. If a product of higher purity is required, the crystalline crop is taken up in a water-containing solution of methanol and is crystallised, optionally under the  
20 addition of activated charcoal. The water-containing solution of methanol has a methanol content of 70 to 99 vol.%, preferably 80 to 99 vol.%, and particularly preferably 90 to 99 vol.%.

The concentration of calcium or magnesium D-pantothenate in  
25 the crystalline product obtained in this way is 60 to 99 wt.%, preferably 70 to 99 wt.%, and particularly preferably 80 to 99 wt.%. The product is suitable for use as a feed additive in animal nutrition.

The concentration of D-pantothenic acid may be determined  
30 by known methods (Velisek; Chromatographic Science 60, 515-560 (1992)). Pure calcium D-pantothenate (> 99 wt.%) has a content of 91.16 wt.% D-pantothenic acid. Pure magnesium D-pantothenate (> 99 wt.%) has a content of 94.73 wt.% D-pantothenic acid.

### Examples

The present invention is described in more detail hereinafter with the aid of examples of implementation.

For this purpose experiments were carried out with the  
5 strain *Escherichia coli* 5069/pFV31 producing D-pantothenic acid, which is registered as FERM-BP 4395 according to the Budapest Agreement at the Fermentation Research Institute, Agency of Industrial Science and Technology in 1-1-3, Higashi, Tsukuba-shi, Ibaraki (Japan) (US-A-5 518 906).

10 Fermentation experiments were also carried out in which the calcium salt of fumaric acid or of aspartic acid was added to the fermentation broth in order to form calcium D-pantothenate directly.

### Example 1

15 Preparation of the calcium salt of D-pantothenate involving the metering in of a calcium hydroxide suspension

#### 1. Preparation of the inoculum (master cell bank)

A sample of *Escherichia coli* FV5069/pFV31 was plated out on LBG agar that had been supplemented with 50 µg per ml  
20 ampicillin. This agar plate culture was incubated for 17 hours at 37°C and then kept in a refrigerated cabinet at +4°C. Selected individual colonies were then propagated further in LBG broth. The LBG broth has the following composition: 10 g/l peptone, 5 g/l yeast extract, 5 g/l  
25 NaCl and 1 g/l glucose. LBG agar contains in addition 12 g/l agar. Ready-for-use preparations may be obtained from Gibco/BRL (Paisley, Scotland, UK) as LB Broth Base or LB agar. The specified media are then obtained by adding 1 g/l glucose. 10 ml cultures that had been obtained in  
30 100 ml Erlenmeyer flasks were incubated for 16 hours at 37°C and 180 rpm in an ESR incubator from Kühner AG (Birsfelden, Switzerland). The cell suspension was then

centrifuged off for 15 minutes at 4000 rpm in a J-6B centrifuge from Beckmann (Hanover, Germany). The cell pellet was resuspended in 10 ml of LBG medium that had been supplemented with 20 % glycerol, was filled in 10 aliquots 5 each of 1 ml [sic] under sterile conditions and frozen at -70°C. These cultures were used as master cell banks.

In order to prepare a working cell bank, LBG medium that had been supplemented with 50 µg/ml ampicillin was added in 10 ml portions to 100 ml Erlenmeyer flasks and then 10 inoculated with 100 µl of the aforescribed master cell bank. The incubation was carried out for 16 hours at 37°C and 180 rpm in an ESR incubator from Kühner AG (Birsfelden, Switzerland).

After the incubation the optical density (OD) of the 15 culture suspension was determined with a LP2W photometer from the Dr. Lange Company (Berlin, Germany) at a measurement wavelength of 660 nm. The optical density was 3.5. The cell suspension was then added under sterile conditions to sterile 30 ml polyethylene test tubes from 20 Greiner (Frickhausen, Germany) and centrifuged at 2500 rpm for 15 minutes in a J-6B type centrifuge from Beckmann (Hanover, Germany). The separated biomass was resuspended in 10 ml of LBG medium that had been supplemented with 20 % glycerol. The cell suspension was next added in 500 µl 25 portions under sterile conditions to 1 ml sterile test tubes from Nalgene (New York, U.S.A.) and frozen at -70°C. The frozen cell suspensions prepared in this way were used as a working cell bank.

## 2. Preparation of a fermentation broth containing calcium 30 D-pantothenate

In order to prepare a fermentation broth containing calcium D-pantothenate the working cell bank was first of all propagated in a shaking flask incubator and was then used to inoculate a preliminary fermenter. The culture from the

preliminary fermenter was used to inoculate the production fermenter.

The SKA medium was used for the shaking flask culture (Table 1). The SKA medium was prepared as follows: 7.0 g  
5  $(\text{NH}_4)_2\text{SO}_4$ , 0.5 g  $\text{KH}_2\text{PO}_4$ , 1.0 g  $\text{K}_2\text{HPO}_4$ , 0.5 g  $\text{MgSO}_4 \cdot 7 \text{H}_2\text{O}$ ,  
0.01 g  $\text{MnSO}_4 \cdot \text{H}_2\text{O}$ , 0.01 g  $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ , 0.005 g  $\text{Fe}_2(\text{SO}_4)_3$  and  
20 g corn steep liquor that had previously been adjusted to  
pH 6.8 with 25 % ammonia solution were weighed out into a  
1 l beaker, which was then made up to 825 g with distilled  
10 water. This salt solution containing corn steep liquor was  
sterilised in an autoclave for 20 minutes at 121°C. A  
solution consisting of 24 g of glucose and 0.002 g of  
thiamine HCl, which had been made up to 125 g with  
distilled water, was sterilised by filtration. 10 g  $\text{CaCO}_3$   
15 were weighed into a 100 ml flask and sterilised in an  
autoclave for 20 minutes at 123°C. The SKA medium was  
obtained by combining the two aforementioned components  
with the salt solution containing the corn steep liquor.

This SKA medium was added in 12.5 ml portions to 100 ml  
20 Erlenmeyer flasks and then inoculated with 0.5 ml of a cell  
suspension. A frozen sample of the working cell culture  
that had been diluted 1:100 with sterile physiological  
saline was used as cell suspension. The incubation was  
carried out for 20 hours at 32°C and 150 rpm in a RC-1-TK  
25 incubator from Infors AG (Bottmingen, Switzerland). The  
optical density subsequently determined at a measurement  
wavelength of 660 nm (OD 660) was 12.5.

In order to inoculate 20 kg of the pre-culture medium A1-  
102 that had been obtained in a 42 l capacity stirred  
30 reactor fermenter from Bioengineering (Wald, Switzerland,  
LP-42 Model), 0.5 ml of the SKA mediums was diluted 1:100  
and 50 ml of the resulting suspension were added to the  
fermenter. The pre-culture medium A1-102 contained the  
constituents listed in Table 2. The culture was cultivated

for 15.5 hours at a temperature of 37°C, a volume-specific aeration of 0.5 vvm, an oxygen partial pressure of 20% of the atmospheric saturation, and a pH of 6.5 until an OD660 of 11.3 had been reached.

- 5 For the inoculation of 5830 g of the principal culture medium M1-425 that had been obtained in 14 l capacity stirred reactor fermenters from B.Braun (BBI, Germany, Melsungen, Biostat E/ED Model), 423 ml of the second pre-culture were added to the medium A1-102. The principal
- 10 culture medium M1-425 contained the constituents listed in Table 3. The culture was first of all cultivated for 6.5 hours at a temperature of 37°C, a volume-specific aeration of 0.75 vvm, a minimum stirring rate of 400 rpm, a pH of 6.5 until an OD660 of 18.6 had been reached, and an oxygen
- 15 partial pressure of 2 % of atmospheric saturation. The culture was then cultivated for a further 41 hours at a temperature of 37°C, an oxygen partial pressure of 2 % of atmospheric saturation and a pH 6.0, until an OD660 of 66.8 had been reached. After a fermentation time of 13 hours  $\beta$ -
- 20 alanine was added in a concentration of 152.7 g in 570 ml H<sub>2</sub>O over a period of 34.5 hours. After a fermentation time of 21.5 hours, a 10% Ca(OH)<sub>2</sub> solution was added over a period of 26 hours in order to stabilise the pH. 3.43 kg of the M2-257 medium having a glucose concentration of
- 25 650.8 g/l and a thiamine HCl concentration of 35.7 mg/l were metered in within 41 hours.

The optical density (OD) was then determined with a LP1W type digital photometer from Dr. Bruno Lange GmbH (Berlin, Germany) at a measurement wavelength of 660 nm, and the

30 concentration of formed D-pantothenic acid was determined by means of HPLC (Hypersil APS 2 5  $\mu$ m, 250x5 mm, RI detection).



A calcium D-pantothenate concentration of 49.8 g/l measured as D-pantothenic acid was determined in the fermentation sample after 47.5 hours.

Table 1

## Composition of the SKA Medium

Component	Concentration (per litre)
Glucose	25 g
Corn Steep Liquor	20 g
$(\text{NH}_4)_2\text{SO}_4$	7 g
$\text{KH}_2\text{PO}_4$	0.5 g
$\text{K}_2\text{HPO}_4$	1 g
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	0.5 g
$\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	5 mg
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	10 mg
$\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$	1 mg
$\text{CaCO}_3$	10 g
Thiamine Chloride · HCl	2 mg
Structol	0.7 g

Table 2

## Composition of the A1-102 Medium

Component	Concentration (per litre)
Glucose	25 g
Corn Steep Liquor	20 g
$(\text{NH}_4)_2\text{SO}_4$	7 g
$\text{KH}_2\text{PO}_4$	0.5 g
$\text{K}_2\text{HPO}_4$	1 g
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	0.5 g
$\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	10 mg
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	10 mg
Thiamine Chloride · HCl	3 mg
Structol	0.6 g

Table 3

Composition of the M1-425 Medium

Component	Concentration (per litre)
Glucose	18 g
Corn Steep Liquor	40 g
$\beta$ -Alanine	15 g
$\text{NH}_4\text{Cl}$	6.8 g
$(\text{NH}_4)_2\text{SO}_4$	3.3 g
$\text{KH}_2\text{PO}_4$	0.6 g
$\text{K}_2\text{HPO}_4$	1.2 g
$\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$	0.67 g
$\text{MnSO}_4 \cdot \text{H}_2\text{O}$	10 mg
Thiamine Chloride $\cdot \text{HCl}$	1.6 mg
Structol	0.6 g

## Example 2

Preparation of the calcium salt of D-pantothenic acid involving the metering in of a calcium acetate solution.

The Escherichia coli strain FV5069/pFV31 was cultivated in  
5 the SKA shaking flask medium as described in Example 1 (Table 1). The suspension was then diluted 1:100 and 17.5 ml of the suspension in this dilution was used to inoculate 7 kg of A1-102 medium (Table 2) in a 14 l capacity stirred reactor fermenter (BBI, Germany,  
10 Melsungen, Biostat E Model).

The culture was cultivated for 15.5 hours at a temperature of 37°C, a volume-specific aeration of 0.5 vvm, an oxygen partial pressure of 20% of atmospheric saturation and a pH of 6.5 until an OD660 of 11.6 had been reached.

15 After inoculation in the production fermenter (14 l stirred reactor fermenter, BBI, Germany, Melsungen, Biostat ED Model) the same cultivation conditions as described in Example 1 were established. A 25 wt.% solution of calcium acetate was used to adjust the pH. The replacement of  
20 ammonia water by calcium acetate was complete after 24 hours. The overall process time of the fermentation was 52 hours. The metering in of 3.4 kg of M2 medium was effected within a period of 40 hours. At the end of the fermentation a calcium D-pantothenate concentration of 43.7 g/l,  
25 measured as D-pantothenic acid, was determined.

## Example 3

Preparation of a product containing calcium D-pantothenate

1.0 l of the fermentation broth prepared according to the method described in Example 1 and containing calcium D-  
30 pantothenate was first of all evaporated in vacuo at 60°C in a rotary evaporator (Büchi Rotavapor RE-120 laboratory rotary evaporator, Büchi-Labortechnik GmbH, Constance,

Germany) to reduce the liquid fraction to about 50% dry content. The broth concentrated in this way was then spray dried to obtain the calcium salt of pantothenic acid (Büchi-190 laboratory spray dryer, inlet temperature 107°C, 5 outlet temperature 78°C, -40 mbar, 600 NL/h, Büchi-Labortechnik GmbH, Constance, Germany).

The product prepared in this way had a content of 42 wt.% measured as D-pantothenic acid.

#### Example 4

#### 10 Preparation of a biomass-free product containing calcium D-pantothenate

The biomass was first of all separated by centrifugation (Biofuge-Stratos laboratory centrifuge, Heraeus, Düsseldorf, Germany; 20 minutes, 4,000 rpm) in 1.0 l of the 15 calcium D-pantothenate-containing fermentation broth prepared according to the method described in Example 1. The broth treated in this way was then processed *in vacuo* at 60°C in a rotary evaporator (Büchi Rotavapor RE-120 laboratory rotary evaporator, Büchi-Labortechnik GmbH, 20 Constance, Germany) to reduce the liquid fraction to about 50 % dry content. The broth concentrated in this way was then spray dried to form the calcium salt of pantothenic acid (Büchi-190 laboratory spray dryer, inlet temperature 107°C, outlet temperature 78°C, -40 mbar, 600 NL/h, Büchi- 25 Labortechnik GmbH, Constance, Germany).

The product prepared as described above had a content of 64 wt.%, measured as pantothenic acid.

#### Example 5

#### Preparation of crystalline calcium D-pantothenate

30 3,600 g of the fermentation broth prepared according to Example 1 and containing calcium D-pantothenate were

separated from the biomass by filtration and the resultant filtrate was reduced to 700 g by evaporation at 60°C in a rotary evaporator (Büchi Rotavapor RE-151 laboratory rotary evaporator, Büchi-Labortechnik GmbH, Constance, Germany).

- 5 The residue was then taken up and dissolved in 3,200 g of methanol. After cooling the solution to room temperature the insoluble constituents were separated by centrifugation (Biofuge-Stratos laboratory centrifuge, Heraeus, Düsseldorf, Germany; 20 minutes, 4,000 rpm). The
- 10 supernatant clarified in this way was concentrated to dryness once more in a rotary evaporator and the residue was taken up again in 950 g of methanol. After cooling to a temperature of 2°C the crystallisation of the calcium salt of pantothenic acid from the concentrated solution
- 15 thereby obtained was started by inoculation with 2.5 g of crystalline calcium D-pantothenate.

After separating the organic phase *in vacuo* a crystalline product was obtained having a content of calcium D-pantothenate of 84 wt.%.

- 20 In a further recrystallisation step the concentration was successfully increased to 96 wt.%.

#### Example 6

Preparation of the magnesium salt of D-pantothenic acid involving the metering in of a magnesium hydroxide

- 25 suspension

- The preparation of the inoculum for the primary culture was carried out as described in Example 1. For the inoculation of 5830 g of the primary culture medium M1-425 that had been obtained in 14 l capacity stirred reactor fermenters
- 30 from B.Braun (BBI, Germany, Melsungen, Biostat E/ED Model) 846 ml of the second pre-culture in medium A1-102 were added. The primary culture medium M1-425 contained the constituents listed in Table 3. The culture was first of

all cultivated for 6.5 hours at a temperature of 37°C, a volume-specific aeration of 0.75 vvm, a minimal stirring rate of 400 rpm and a pH of 6.5 until an OD660 of 22.0 had been reached, and an oxygen partial pressure of 2 % of atmospheric saturation. The culture was then cultivated for a further 48 hours at a temperature of 37°C, an oxygen partial pressure of 2 % of atmospheric saturation, and a pH of 6.0 until an OD660 of 67.6 had been reached. After a fermentation time of 23.0 hours, a 15 % Mg(OH)<sub>2</sub> suspension was metered in over a period of 31.5 hours to stabilise the pH. 4.28 kg of the medium M2-261 having a glucose concentration of 584.7 g/l, a β-alanine concentration of 50.7 g/l and a thiamine HCl concentration of 35.7 mg/l were metered in within 48 hours.

The optical density (OD) was then measured with an LP1W type digital photometer from Dr. Bruno Lange GmbH (Berlin, Germany) at a measurement wavelength of 660 nm and the concentration of formed D-pantothenic acid was determined by means of HPLC (Hypersil APS 2 5 µm, 250x5 mm, RI-Detection).

A magnesium D-pantothenate concentration of 48.2 g/l measured as D-pantothenic acid was determined in the fermentation end sample after 54.5 hours.



**Patent Claims**

1. Process for preparing alkaline earth salts of D-pantothenic acid or mixtures containing the latter, from fermentation broths,  
5 characterised in that
  - a) the fermentation is carried out in the presence of alkaline earth compounds
  - b) after completion of the fermentation the biomass is optionally removed in whole or in part,
  - 10 c) the thus prepared fermentation broth is concentrated, and
  - d) the alkaline earth salt or salts of D-pantothenic acid is/are obtained therefrom in pure form or as a mixture that contains the constituents of the  
15 fermentation broth.
2. Process according to claim 1,  
characterised in that  
the commencement of the addition of the alkaline earth  
20 compound begins when the concentration of D-pantothenic acid is 0.5 to 70 g/l, preferably 5 to 35 g/l.
3. Process according to claim 1,  
characterised in that  
the alkaline earth compound is added to the  
25 fermentation broth in a molar ratio of 1 : 0.5 to 1 : 20, in particular 1 : 1.3 to 1 : 10, relative to D-pantothenic acid.
4. Process according to claim 3,  
characterised in that  
30 alkaline earth-containing compounds are added during the fermentation to the fermentation broth, batchwise

or continuously, individually or in a mixture, at least in the same stoichiometric ratio as D-pantothenic acid is formed.

5. Process according to claims 1, 2 or 3,  
5 characterised in that  
a calcium salt of one or more organic acids, in particular of fumaric acid or aspartic acid, is added.
6. Process according to claims 1, 2 or 3,  
characterised in that  
10 calcium hydroxide, calcium oxide, magnesium oxide or magnesium hydroxide are used as alkaline earth compounds.
7. Process according to claim 1,  
characterised in that  
15 a) the biomass is separated from the fermentation broth containing the alkaline earth salts of D-pantothenic acid,  
b) the cell-free fermentation broth is concentrated,  
c) a hydrophilic organic solvent is added to the  
20 concentrate thus obtained, and then  
d) the alkaline earth salts of pantothenic acid are crystallised and if necessary purified further.
8. Process according to one or more of the preceding  
claims,  
25 characterised in that  
one or more various alkaline earth salt(s) of D-pantothenic acid is/are added to the constituents of the fermentation broth and mixture containing one or more alkaline earth salts of D-pantothenic acid.

## INTERNATIONAL SEARCH REPORT

International Application No  
PCT/EP 01/04651

**A. CLASSIFICATION OF SUBJECT MATTER**  
 IPC 7 C12P13/02

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the International search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, BIOSIS, CHEM ABS Data

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 493 060 A (TAKEDA CHEM. IND., LTD.) 1 July 1992 (1992-07-01) cited in the application page 2; examples 6,7	1-8
X	EP 0 590 857 A (TAKEDA CHEM. IND., LTD.) 6 April 1994 (1994-04-06) cited in the application page 10-11	1-8
X	WO 97 10340 A (TAKEDA CHEM. IND., LTD.) 20 March 1997 (1997-03-20) cited in the application see especially p. 15 and 21	1-8
X	GB 683 423 A (FISONS LTD.) 26 November 1952 (1952-11-26) page 2	8

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

## \* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the International filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the International filing date but later than the priority date claimed

- "T" later document published after the International filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "&" document member of the same patent family

Date of the actual completion of the International search

8 August 2001

Date of mailing of the International search report

20/08/2001

Name and mailing address of the ISA

European Patent Office, P.B. 5618 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,  
Fax: (+31-70) 340-3016

Authorized officer

Douschan, K

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/EP 01/04651

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP 0493060 A	01-07-1992	JP 3109884 B JP 5023191 A	20-11-2000 02-02-1993
EP 0590857 A	06-04-1994	CN 1095105 A,B CN 1225394 A CN 1225388 A CN 1225392 A CN 1225358 A JP 6261772 A US 5518906 A	16-11-1994 11-08-1999 11-08-1999 11-08-1999 11-08-1999 20-09-1994 21-05-1996
WO 9710340 A	20-03-1997	AU 6943696 A CN 1201491 A EP 0859848 A JP 9135687 A SK 30598 A US 5932457 A	01-04-1997 09-12-1998 26-08-1998 27-05-1997 11-01-1999 03-08-1999
GB 683423 A	26-11-1952	NONE	